# Effect of Sodium Ascorbate or Alpha-Tocopherol on the Resin-Dentin Interface and Bond Strength after Endodontic Treatment and Bleaching

Efecto del Ascorbato de Sodio o Alfa-Tocoferol sobre la Interfaz Resina-Dentina y la Fuerza de Unión Después del Tratamiento de Endodoncia y Blanqueamiento

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**ABSTRACT:** The objective of this in vitro study was to assess the effects of two antioxidants (sodium ascorbate [SA] and alpha-tocopherol [AT]) on the adhesive interface and dentin bond strength immediately after bleaching with 38 % hydrogen peroxide (38HP) in endodontically-treated teeth. Two stages of experimentation were carried out. Bovine incisors were allocated into four groups (n = 10/group for each experiment): NB, non-bleached restored crowns; 38HP, bleached and immediately restored crowns; 38HP-SA, bleached crowns and SA use; and 38HP-AT, bleached crowns and AT use. Hybrid layer length in dentin ( $\mu$ m) and bond strength (MPa) were assessed with confocal microscopy laser and micro-shear bond strength ( $\mu$ SBS) test, respectively. Failure mode was determined by stereomicroscope. Data analysis was performed with analysis of variance (ANOVA) and Kruskal-Wallis, Dunn, Tukey, and Fisher-exact tests (a = 0.05). Higher values of hybrid layer length were observed similarly in the NB and 38HP-SA groups. The highest  $\mu$ SBS mean values were observed in the NB group (18.51 ± 1.33), whereas the SBS values for 38HP-AT (1.68 ± 0.32) were similar to the 38HP group (1.61 ± 0.51) (p > 0.05) and significantly lower than the 38HP-SA group (5.78 ± 0.71). Adhesive failures were predominant in the 38HP and 38HP-AT groups. Cohesive and mixed failures were mostly observed in the NB and 38HP-SA groups, respectively. In conclusion, AT has no immediate effect on the hybrid layer formation and  $\mu$ SBS of dentin. Although SA promotes an increase in hybrid layer formation, it was not reflected in the  $\mu$ SBS values.

KEY WORDS: bleaching, alpha-tocopherol, sodium ascorbate, bond strength, hybrid layer.

### INTRODUCTION

Dental bleaching is one of the non-invasive procedures that meets the public demand for cosmetic dentistry (Carey, 2014). During bleaching, free radicals formed by the interaction of bleaching agent with the tooth structure, act by reducing the chromophore molecules and thus making them less pigmented (Elawsya *et al.*, 2021). In some cases, where the color changes have not sufficiently resolved with bleaching or there is a significant loss of tooth structure, restorative procedures are necessary on the whitened structure (Cavalli *et al.*, 2018). However, residual peroxides can adversely affect the bond strength of adhesive restorations to tooth structure (Cvitko *et al.*, 1991).

A recent meta-analysis (Imani *et al.*, 2020) indicated that shear bond strength to enamel is negatively affected by bleaching procedures. Hence, post bleaching, the current recommended wait time ranges from 24 h to 28 days, to perform definitive restorations (Cavalli *et al.*, 2001; Unlu *et al.*, 2008; Comlekoglu *et al.*, 2010). However, in endodontically treated teeth, this is not always feasible. In addition to the cost and time of new clinical sessions and

Department of Restorative Dentistry, Araraquara Dental School, São Paulo State University (UNESP), Araraquara, Brazil. This study was financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Brazil - Finance Code 001. inconvenience to the patient, a long elapsed time to the restorative treatment presents with risk of coronary failure, microleakage of temporary restorations, and recurrence of discoloration of recently bleached teeth (Carvalho *et al.*, 2018). This is a frequent problem encountered by dental surgeons in clinical practice, and the absence of a consensual protocol has hindered the basis for making clinical decisions.

Use of antioxidants immediately after dental bleaching is one promising alternative for achievement of the adhesive procedure (Garcia et al., 2012). Ascorbic acid is an effective antioxidant, and its low toxicity indicates that the biological risks of its use are minimal. However, ascorbic acid has a low pH (around 4.0) that could cause over-conditioning of the dental surface (Lai et al., 2001). Therefore, effects of sodium ascorbate (SA), which has a neutral pH (around 7), have been studied to reverse the negative impact of bleaching on bonding procedure of adhesive restorations (Freire et al., 2009; Garcia et al.). Alphatocopherol (AT) is considered the most effective fatsoluble antioxidant available in nature (Burton & Ingold, 1989) and has beneficial effects on various health conditions caused by free radicals in organic systems (Cross et al., 1987). Theoretically, this antioxidant would be able to trap free radicals in the bleached surface without converting them into reactive intermediates that could compromise adhesive restorative procedures (Sasaki et al., 2009).

A recent in vitro study demonstrated the nonsignificant short-term effect of AT in reversing the impairment of bond strength in endodontically-treated tooth bleached with sodium perborate (Harrison et al., 2019). However, bleaching procedures on endodontically treated teeth can be challenging and require more effective treatment (Doumani et al., 2019). Consequently, hydrogen peroxide has been widely used in these cases (Souza-Gabriel et al., 2011). Higher concentrations of hydrogen peroxide are associated with significant reduction in enamel shear bond strength. Furthermore, the application time was pointed out as an important factor for damage to the enamel surface (Imani et al.). The effects of dental bleaching with hydrogen peroxide associated with or without the use of antioxidants in immediate adhesive restorative procedures on dentin of endodontically treated teeth have been less explored and remain unclear.

Thus, this *in vitro* study aimed to assess the effects of two antioxidants (SA and AT) on the performance of adhesive restorative procedures performed

immediately after simulated bleaching with 38 % hydrogen peroxide in endodontically-treated teeth. The hypotheses that the use of these antioxidants would reverse the negative effect of the bleaching procedure on (H1) the hybrid layer formation and (H2) the microshear bond strength ( $\mu$ SBS), were tested. Furthermore, the effects of antioxidants on the adhesive interface were qualitatively assessed.

# MATERIAL AND METHOD

**Experimental design and ethical aspects.** This *in vitro* study was conducted in two stages of experimentation: effect of antioxidants on the dentinadhesive interface (Experiment 1) and on the microshear bond strength (mSBS) (Experiment 2) on the dentin of endodontically treated teeth subjected to bleaching.

According to the study protocol, four groups were composed:

**1)** Non-bleached + Restored - positive control (NB): The specimens were etched, adhesive system (Scothbond Multi-Purpose; 3M ESPE, St. Paul, MN, USA) was applied, and the dental crowns were restored with composite resin (Filtek Z-250; 3M, St. Paul, MN, EUA). The primer from the adhesive system was applied onto the dentin using a brush tip (KG Sorensen, São Paulo, SP, BR) and gently dried for 5 s. After 5 s, dentin adhesive was applied and light-cured with a lightemitting diode (LED)-laser system (LED Bluephase; IvoclarVivadent, Schan, Liechtenstein, AL) with an intensity of 1,200 mW/cm2 for 10 s. The pulp chamber was then restored with composite resin, with 2 mm thick increments and light-cured for 40 s at each increment.

2) Bleached with 38 % hydrogen peroxide + Restored without antioxidant use - negative control (38HP): The buccal surface and pulp chamber were subjected to bleaching with 38 % hydrogen peroxide (Opalescence Xtra Boost; Ultradent Products Inc, South Jordan, UT, USA). The bleaching gel was handled according to the manufacturer's recommendations and applied at about  $\pm$  2.0 mm thickness to the buccal surface of the dental crown and within the pulp chamber. The gel was applied for 15 min, aspirated, and applied twice under similar conditions described above. Then the dental crown was copiously rinsed with gentle water-spray for 1 min, dried and immediately restored, similar to group NB. **3)** Bleached with 38 % hydrogen peroxide + Restored after 10 % SA gel use (38HP-SA): After the bleaching protocol similar to 38HP group, 10 % SA gel (Apothicário, Araçatuba, SP BR) was applied for 15 min on the buccal surface and 10 min within the pulp chamber. The antioxidant was removed with a gently water-spray for 15 s, and the tooth was immediately restored similar to the 38HP group.

4) Bleached with 38 % hydrogen peroxide + Restored after 10 % AT gel use (38HP-AT): Similar procedures to 38HP-SA group were performed, but using 10 % AT gel (Apothicário, Araçatuba, SP BR) as an antioxidant. After 24 h, the dental crowns were crosssectioned from the root portion at the cemento-enamel junction and longitudinally sectioned in bucco-lingual direction using a hard tissue cutting machine (Isomet; Buehler, Lake Bluff, IL). The distal section from each dental crown was selected, and the dentin-restoration interface was flattened with #600 followed by #1200 wet sandpaper (Norton, Lorena, SP, BR), under running water-cooled circular polisher (Arotec, Cotia, SP, BR).

The study protocol was approved by the local Ethics Committee.

**Sample size calculation.** For the sample size calculation was considered the dentin tags penetration and bond strength values of a previous study (Harrison *et al.*). Using a significance level of 0.05 and a test power of 80 %, the sample size necessary was 10 specimens. Considering all variables of the 2 experiments adopted in the study, 80 bovine teeth were selected.

**Specimens preparation.** Forty bovine incisors with similar crown and root anatomy (radiographically confirmed) were stored in thymol solution (0.1 %; pH 7.0) at 4°C until the beginning of the research. The pulp chamber access was performed with a 1014 spherical diamond drill (KG Sorensen, Cotia, SP, Brazil), and the access diameter was standardized with a #12 steel spherical drill.

Root canals were instrumented by crown-down technique (Morgan & Montgomery, 1984), 1 mm from the root apex, up to #80 K-file, irrigated with 5 mL of 2.5 % sodium hypochlorite at each file change. After the biomechanical preparation, the root canals were irrigated with 3 mL of 17 % EDTA (Biodinâmica, Ibiporã, PR, Brasil) for 3 min, and 10 mL of distilled water. The canals were then aspirated and dried with absorbent paper points (Dentsply-Herpo, Petrópolis, RJ, Brasil).

The root canals were obturated using gutta-percha (Dentsply Ind Com Ltda, Petrópolis, RJ, Brasil) and epoxy-based sealer (AH Plus; Dentsply De Trey, Konstanz, Germany). The gutta-percha was removed 3 mm below the tooth cervical line, and a cervical barrier with self-curing glass ionomer (Maxxion R A3; FGM ProdutosOdontológicos Ltda., Joinville, SC, Brasil) was placed at the cement-enamel junction.

The access cavity was restored with provisional restorative cement (IRM; Dentsply Ind Com Ltda, Petrópolis, RJ, Brasil), and the teeth were immediately immersed in artificial saliva (Faculdade de CiênciasFarmacêuticas de Ribeirão Preto-USP, Ribeirão Preto, SP, Brasil). After 24 h, the temporary restoration was removed, and the pulp chamber was irrigated with 2.5 mL of 2.5 % sodium hypochlorite, and 2.5 mL of distilled water. Thirty-seven percent phosphoric acid (Condac 37; FGM Produtos Odontológicos Ltda., Joinville, SC, Brasil) was used to etch the dentin for 15 s and was rinsed off with distilled water (Kuga *et al.*, 2012).

After 24h of the experimental procedures, the dental crowns were cross-sectioned from the root portion at the cemento-enamel junction and longitudinally sectioned in bucco-lingual direction using a hard tissue cutting machine (Isomet; Buehler, Lake Bluff, IL). The distal section from each dental crown was selected, and the dentin-restoration interface was flattened with #600 followed by #1200 wet sandpaper (Norton, Lorena, SP, BR), under running water-cooled circular polisher (Arotec, Cotia, SP, BR).

The specimens were washed with distilled water, and the surface was polished with aluminium oxide (Arotec, São Paulo, SP, BR) at 30 mm granulation, and felt disc at circular polisher. Then, the specimens were immersed in distilled water and stirred in an ultrasonic tank (Cristófoli, Campo Mourão, PR, USA) for 10 min.

The specimens were dried with absorbent paper, and the whole surface was etched with 37 % phosphoric acid (Condac 37; FGM Produtos Odontológicos Ltda., Joinville, SC, Brasil) for 60 s. The surfaces were rinsed with 50 mL of distilled water, dehydrated with air-spray, and individually fixed on a glass slide with the etched surface kept at horizontal position.

For the  $\mu$ SBS test, otherforty bovine incisors were obtained and preserved similar to the previous experiment. The crowns were transversely sectioned at the cement-enamel junction and the roots were

discarded. The crown was sectioned in the mesiodistal direction using a hard tissue cutting machine (Isomet 100, Buehler, Lake Bluff, IL) obtaining 10 mm length slabs. The buccal surface of the slabs was ground with a polishing machine (DP-10; Panambra, Struers, Ballerup, DI) using #180 silicon carbide sandpapers in order to obtain the dentin exposure and make it flat.

The exposed dentin was polished with #320 and #6000 sandpapers for 20 s. The slabs were then embedded into polystyrene matrix molds (16.5 mm width  $\pm$  25.0 mm length), with acrylic resin (Classic Jet, São Paulo, SP, BR) leaving the dentin surface uncovered by the acrylic resin. After the acrylic resin polymerization, the slabs were randomly allocated into 4 groups (n = 10), similar to Experiment 1.

After all the dentin treatment protocols, four cylinders made of composite resin specimens were prepared on the buccal surface: two at mesial and two at distal. A Tygon tube transparent matrix (Tygon tube, R-3603, Saint-Gobain Performance Plastics, Maime Lakes, FL, USA) with 0.72 mm internal diameter and 1.0 mm height was used for the composite resin filling. (Filtek Z-250; 3M, St. Paul, MN, USA).

Hybrid-layer length in dentin. Each specimen was analyzed with a laser confocal microscope (LEXT OLS4100; Olympus, Shinjuku-ku, Tokyo, JP), using specific software (Olympus Stream; Olympus, Shinjuku-ku, Tokyo, JP), at a magnification of 1024¥ (Guarda et al., 2020). The images were saved as TIFF format, and the hybrid layer formation length in dentin was measured using the Image J program, calibrated in micrometers (mm). The intra dentin length from the formed hybrid layer was measured in 100 mm, from the buccal surface of the middle-third of the dental crown. The measurement was performed for every 10 µm, and a total of 10 analyses were obtained for each specimen. The arithmetic average from these measurements indicated the hybrid layer formation in each specimen.

Micro-shear bond strength ( $\mu$ SBS) test. After obtaining the specimens, they were stored in a humid environment and mSBS test was performed after 24 h. All slabs were fixed inside a metal matrix so that the composite cylinder specimens were placed perpendicularly to the load cell of 500 N.

An orthodontic wire (0.2 mm diameter) held the composite cylinder base, and all the specimens were subjected to compressive loading in an

electromechanical testing machine (DL2000; EMIC, Pinhais, PR, Brazil) with a crosshead speed of 0.5 mm/ min until the end of mSBS test (Shimada *et al.*, 2003). The bond strength was obtained from the maximum force (N) divided by the union area (mm<sup>2</sup>), in MPa. The arithmetic average was calculated for the four specimens from each slab and called an average specimen.

**Failure mode analysis.** After the mSBS test, failure modes were determined analyzing a failed surface of each sample with a stereomicroscope (SZ - PT; Olympus, Japan; 40X magnification). The failures were classified as: A) adhesive, if the failure occurred totally at the adhesive interface, B) cohesive, if the failure was observed exclusively in dentin or resin, or C) mixed when a combination of adhesive and cohesive failures was observed (Sasaki *et al.*). Furthermore, representative images of failure modes were obtained with Scanning Electron Microscopy (SEM), at 500x magnification.

**Statistical Analysis.** The non-normal data distribution was verified with the Shapiro-Wilk test for the hybridlayer formation data. Thus, Kruskal-Wallis and Dunn tests were applied. Confocal fluorescence images provided qualitative data regarding the adhesive interface. The bond strength data were analyzed using one-way ANOVA and Tukey test. The failure modes data were analyzed using Fisher's exact test. Level significance at 5 % was adopted for all analyses. Data acquired from SEM provided qualitative data regarding the failure modes.

# RESULTS

Table I presents the hybrid layer formation values ( $\mu$ m). The 38HP-SA group presented a hybrid layer formation similar to the NB group. With the median values of hybrid layer formation significantly inferior, the 38HP and 38HP-AT groups did not differ from each other (p> 0.05). Confocal fluorescence images (Fig. 1) showed that the hybrid layer formation was more uniform in the NB and 38HP-SA groups.

The highest  $\mu$ SBS mean values (in MPa) (Table II) were observed in the NB group (18.51 ± 1.33). In a decreasing sequence, the 38HP-AS (5.78±0.71) group presented higher bond strength than the 38HP-AT (1.68 ± 0.32) and 38HP (1.61±0.51) groups, whose values were similar to each other (p> 0.05).

Table I. Median, maximum, and minimum values, and the first and third quartile of the intra-dentin length from the hybrid layer formation ( $\mu$ m).

	NB	38HP	38HP-SA	38HP-AT
Median	11.23 <sup>ª</sup>	3.63 <sup>D</sup>	9.86 <sup>a</sup>	3.94 <sup>b</sup>
Max-Min	15.26-9.91	4.86-0.30	10.60-8.63	5.13-3.22
1Q-3Q	10.26-11.65	1.55-3.98	9.06-10.24	3.47-4.48

a,b Different letters indicate significant statistical difference (Dunn test, p<0.05). Max: maximum value and Min: minimum value; 1Q: first quartile and 3Q: third quartile. NB - restored crowns, 38HP - bleached and immediately restored crowns, 38HP-SA - bleached crowns and sodium ascorbate use, and 38HP-AT - bleached crowns and alpha-tocopherol use.



Fig. 1. Representative images showing a hybrid layer formation analysis for all groups obtained by confocal laser microscopy (1024¥ magnification). A - Restored crowns (NB), B - bleached and immediately restored crowns (38HP), C - bleached crowns and sodium ascorbate use (38HP-SA), D - bleached crowns and alpha-tocopherol use (38HP-AT).

Table II. Micro-shear bond strength (in N	MPa) of the adhesive
system to dentin	

	NB	38HP	38HP-SA	38HP-AT
Mean	18.51 <sup>a</sup>	1.61 <sup>°</sup>	5.78 <sup>⁰</sup>	1.68 <sup>°</sup>
SD	1.33	0.51	0.71	0.32

a,b Different letters indicate significant statistical difference (Tukey test, p<0.05). SD - standard deviation, NB - restored crowns, 38HP - bleached and immediately restored crowns, 38HP-SA - bleached crowns and sodium ascorbate use, and 38HP-AT - bleached crowns and alpha-tocopherol use.

The failure modes showed in Figure 2 presented significant differences between groups (p<0.001). Cohesive failures were predominant in the NB group (67.5 %). Adhesive failures (83 %) were mostly observed in the 38HP group similar to that of the 38HP-AT group. Furthermore, in the 38HP-SA group, mixed failures were predominant (72.5 %). SEM representative images of each failure mode are shown in Figure 3.





# % Failure mode

Fig. 2. Failure mode analysis for all groups after the micro-shear bond strength test. A significant statistical difference is observed (Fisher-exact test, p<0.001). NB - restored crowns, 38HP - bleached and immediately restored crowns, 38HP-SA - bleached crowns and sodium ascorbate use, and 38HP-AT - bleached crowns and alpha-tocopherol use



Fig. 3. Representative images showing (A) adhesive, (B) mixed, and (C) cohesive failures analyzed by scanning electronic microscopy (500¥ magnification).

### DISCUSSION

The antioxidants evaluated in the present study demonstrated distinct immediate effects on the tooth structure subjected to bleaching with 38 % hydrogen peroxide. SA reversed the negative effect of bleaching on the formation of a hybrid layer, presenting results similar to the non-bleached tooth structure (NB group). However, the compromise in hybrid layer formation was similar in 38HP-AT group and tooth structure without use of antioxidant post-bleaching (38HP group). Thus, H1 was rejected.

Dentinal tubules are permeable to bleaching agents, thus representing reservoirs of oxygen free radicals (Perdigão, 2010). These residual peroxides

releasing free radicals from the bleaching agents persist in the tooth structure until they are removed by pulp microcirculation and/or diffusion from the external surface. Based on a reduced pulp microcirculation, greater external diffusion is expected. In endodontically treated teeth, a clinical condition simulated in this study, the accumulation of free radicals is very critical. Thus, increased levels of peroxide may be present at the adhesive interface, compromising resinous materials infiltration in the tubules and the formation of a uniform hybrid layer (Abraham *et al.*, 2013). A previous study has shown that compared with the non-bleached tooth surface, bleaching causes approximately two-thirds reduction in the penetrability of resin tags on the tooth surface (Titley *et al.*, 1991). This can justify the results of the present study, where the 38HP group presented a hybrid layer formation significantly inferior to the NB group.

Our findings demonstrated that the application of 10 % SA immediately after bleaching, allowed the adhesive procedure to provide a hybrid layer similar to the NB group. This finding suggests that similar the occurrence in bleached enamel, SA is effective in eliminating peroxide free radicals, allowing faster polymerization of adhesive materials without any premature interruption (Briso *et al.*, 2012; Thapa *et al.*, 2013; Briso *et al.*, 2014; Kavitha *et al.*, 2016; Elawsya *et al.*). However, no differences in the hybrid layer formation were observed between the 38HP-AT and 38HP groups, demonstrating that AT did not have significant effects in reversing the effect of the bleaching procedure on the hybrid layer formation (Thapa *et al.*).

Sasaki *et al.* observed that AT was effective in neutralizing free radicals resulting from bleaching with carbamide peroxide. This may justify the divergence with the results of our study because the impairment of adhesive procedures on a recently bleached surface appears to be directly proportional to the concentration of the bleaching agent (Imani *et al.*). In this study, where a stronger bleaching agent was used (38 % hydrogen peroxide), possibly a higher concentration of the AT gel would be needed to neutralize the accumulation of oxygen in the dentin that probably compromised the formation of the hybrid layer. Thus, it is strongly recommended that the effects of this antioxidant in other concentrations and/or application times be investigated.

In this study, the  $\mu$ SBS test was chosen to assess the adhesive strength. This test has the advantage of greater control of the test area due to the use of tubes of known diameter (Shimada *et al.*). The bleaching procedure, with or without the application of antioxidants, significantly compromised the  $\mu$ SBS compared with the NB group. In addition to creating a reservoir of residual oxygen, bleaching changes the mineral and protein content of dentin (Perdigão). The ineffectiveness of the antioxidants evaluated in the present study in reversing the impairment of bond strength may be associated with this fact. Thus, the H2 was also rejected.

Although application of SA showed an increase in bond strength compared with the bleached surface, its value was significantly lower than that of the nonbleached surface. This result is in agreement with previous studies (Sasaki *et al.*; Nascimento *et al.*, 2019; Zhang *et al.*, 2020), and the primary theory that justifies this finding is that SA can react with hydroxyl crystals or peroxiapatite resulting from bleaching (Briso *et al.*, 2014; Rezaei *et al.*, 2019). Thus, application of 10 % SA gel for 10 min can increase the bond strength to the bleached tooth structure. However, the SBS values were lower than the non-bleached surface. As with enamel, the application of SA can cause overconditioning of the whitened surface (Muraguchi *et al.*, 2007). This can compromise the already delicate adhesion to dentin, justifying these results.

In contrast with the previous studies (Whang & Shin, 2015; Kavitha *et al.*, 2016; Nari-Ratih & Widyastuti, 2019), the findings of this study demonstrated that SA effects were significantly higher than the group using AT. However, in these studies, AT was applied as an alcohol-based solution. Therefore, a similar effect of AT cannot be attributed exclusively to the antioxidant because ethanol has the recognized effect of reversing the compromised bond strength to the whitened surface (Sung *et al.*, 1999). These methodological differences may justify the disagreement with our results.

The µSBS values obtained in this study correlate with the results of the failure analysis. In the 38HP-AT and 38HP groups, adhesive failures were the most frequent. This failure mode, which occurred entirely at the adhesive interface, demonstrates the compromise potential of residual peroxides from bleaching on the adhesive restorative procedures (Cavalli et al., 2018). Furthermore, these results corroborate the non-efficacy of the AT use with respect to hybrid layer formation and bond strength (Thapa et al.; Harrison et al.). The failure mode, predominantly mixed, associated with SA use indicated the benefits of this antioxidant in the adhesive interface compared to the bleached surface with or without the use of AT. These (mixed) failures are attributed to thermo-mechanical ablation that can compromise the physical characteristics of the dentin substrate and the restorative material (Jacker-Guhr et al., 2019). Not surprisingly, in the NB group, the cohesive failure mode was predominant. This finding suggests the improvement of adhesive procedures, when performed without competing with the residual products of bleaching (Kum et al., 2004).

Our study demonstrated that there are no beneficial effects of using 10 % AT gel for 10 min in reversing the negative effects of tooth whitening on adhesive

restorative procedures of endodontically treated teeth. These findings point out that at the moment, in this concentration, application period, and form of presentation, AT is still clinically unviable. More efforts are needed to completely clarify its mechanism and explore its possible correct indication in the adhesive procedures immediately after bleaching.

## CONCLUSION

Use of10 % AT gel had no immediate beneficial effect in the hybrid layer formation and micro-shear bond strength. SA promotes a beneficial effect in hybrid layer formation, which was not reflected on the  $\mu$ SBS. The findings of this study do not suggest any change in the recommended delay time for definitive restorations of the bleached tooth structure.

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RESUMEN: El objetivo de este estudio in vitro fue evaluar los efectos de dos antioxidantes (ascorbato de sodio [AS] y alfa-tocoferol [AT]) sobre la interfaz adhesiva y la fuerza de unión de la dentina inmediatamente después del blanqueamiento con peróxido de hidrógeno al 38 % (38HP) en endodoncia. -Dientes tratados. Se llevaron a cabo dos etapas de experimentación. Los incisivos bovinos se dividieron en cuatro grupos (n = 10 / grupo para cada experimento): NB, coronas restauradas no blangueadas; 38HP, coronas blanqueadas y restauradas inmediatamente; 38HP-AS, coronas blanqueadas y uso SA; y 38HP-AT, coronas blanqueadas y uso de AT. La longitud de la capa híbrida en dentina ( $\mu$ m) y la fuerza de unión (MPa) se evaluaron con láser de microscopía confocal y la prueba de fuerza de unión por micro-cizallamiento (µSBS), respectivamente. El modo de falla se determinó mediante estereomicroscopio. El análisis de los datos se realizó con análisis de varianza (ANOVA) y pruebas de Kruskal-Wallis, Dunn, Tukey y Fisher ( $\alpha$  = 0,05). De manera similar, se observaron valores más altos de longitud de capa híbrida en los grupos NB y 38HP-AS. Los valores medios más altos de µSBS se observaron en el grupo NB (18,51 ± 1,33), mientras que los valores de SBS para 38HP-AT (1,68 ± 0,32) fueron similares a los del grupo 38HP (1,61 ± 0,51) (p> 0,05) y significativamente más bajos que el grupo 38HP-AS (5,78 ± 0,71). Las fallas adhesivas fueron predominantes en los grupos de 38HP y 38HP-AT. Las fallas cohesivas y mixtas se observaron principalmente en los grupos NB y 38HP-AS, respectivamente. En conclusión, la AT no tiene un efecto inmediato sobre la formación de la capa híbrida y el µSBS de dentina. Aunque AS promueve un aumento en la formación de capas híbridas, no se refleja en los valores de µSBS.

PALABRAS CLAVE: blanqueo, alfa-tocoferol, ascorbato de sodio, fuerza de unión, capa híbrida.

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